



Universitat de Lleida

Document downloaded from:

<http://hdl.handle.net/10459.1/64985>

The final publication is available at:

<https://doi.org/10.1111/rda.12937>

Copyright

(c) Blackwell Verlag GmbH, 2017

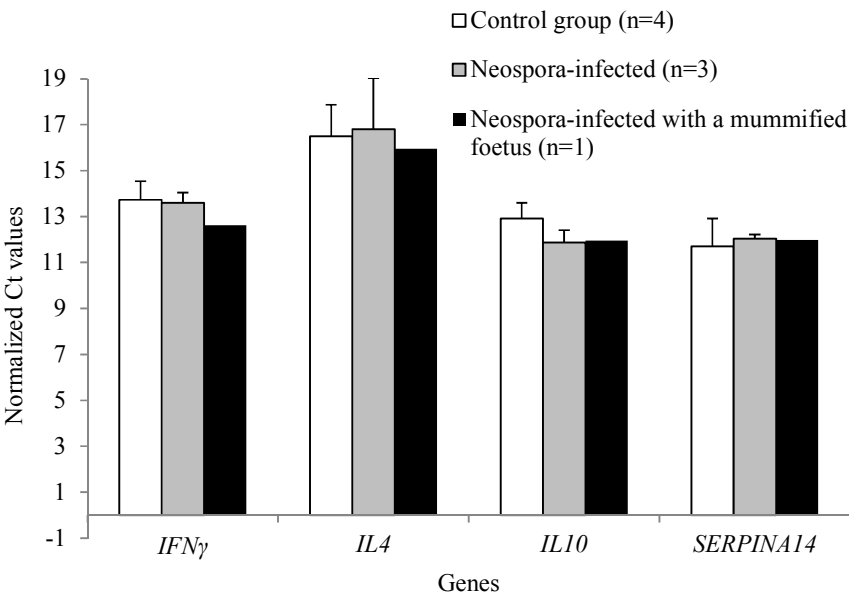
Reproduction in Domestic Animals

Uterine serpin (SERPINA 14) correlates negatively with cytokine production at the foetal-maternal interface but not in the corpus luteum in pregnant dairy heifers experimentally infected with *Neospora caninum*

Journal:	<i>Reproduction in Domestic Animals</i>
Manuscript ID	Draft
Manuscript Type:	Original Article
Date Submitted by the Author:	n/a
Complete List of Authors:	Serrano, Beatriz; University of Lleida, Animal Production Almeria, Sonia; Autonomous University of Barcelona, Mur-Navales, Ramón; University of Lleida, Animal Production López-Helguera, Irene; UdL, García-Ispuerto, Irina; University of Lleida, Animal Production; ALABART, JOSE LUIS; Centro de Investigación y Tecnología Agroalimentaria (CITA) de Aragón, Tecnología en Producción Animal Darwich, Laila; Autonomous University of Barcelona, Animal health and anatomy López-Gatius, Fernando; University of Lleida,
Subject Area:	Embryo/fetus < General reproduction, Immunology < General reproduction, Placenta < General reproduction, cattle < Species:

SCHOLARONE™
Manuscripts

Figure 1



Uterine serpin (SERPINA 14) correlates negatively with cytokine production at the foetal-maternal interface but not in the corpus luteum in pregnant dairy heifers experimentally infected with *Neospora caninum*

Running title: Serpins and cytokines in *Neospora*-infected heifers

B. Serrano-Pérez^{1,2}, S. Almería^{3,4}, R. Mur-Novales¹, I. López-Helguera^{1,2}, I. Garcia-Isperto^{1,2}, J.L. Alabart⁵, L. Darwich^{3,4}, F. López-Gatius^{2,6*}

¹Department of Animal Science, University of Lleida, Spain.

²Agrotecnio Centre, University of Lleida, Spain.

³Centre de Recerca en Sanitat Animal (CReSA) - Institut de Recerca i Tecnologia Agroalimentàries (IRTA), Campus UAB, 08193 Bellaterra, Barcelona, Spain

⁴Departament de Sanitat i Anatomia Animals, Universitat Autònoma de Barcelona (UAB), 08193 Bellaterra, Barcelona, Spain

⁵Instituto Agroalimentario de Aragón - IA2 - (CITA-Universidad de Zaragoza), Zaragoza, Spain

⁶Transfer in Bovine Reproduction SLu, 22300 Barbastro, Spain

*Correspondence, F. López-Gatius. Email address: lopezgatiusf@gmail.com

24 Contents

25

26 *Neospora caninum* is a major cause of abortion in cattle worldwide. However, immune-
27 endocrine interactions during pregnancy in *Neospora*-infected cows remain largely
28 unknown. This study examines gene expression patterns in dairy heifers experimentally
29 infected with *N. caninum* during the second trimester of pregnancy that did not abort (n:
30 4 non-infected control dams and 4 infected dams). Based on the patterns observed,
31 relationships were determined between gamma interferon (*IFN* γ), (Th1 pro-
32 inflammatory cytokine), interleukin 4 (*IL4*) (Th2 pro-gestation cytokine) or interleukin
33 10 (*IL10*) (T regulatory cytokine) and the serine peptidase inhibitor *SERPINA14* in
34 intercaruncular, placental, uterine lymph node (UTLN) and luteal tissue samples.
35 Intercaruncular *SERPINA14* expression was negatively correlated with *IFN* γ expression
36 in cotyledon samples (sr: -0.943; P= 0.005) and with *IL4* expression in UTLN (sr: -
37 0.886; P= 0.019). A trend towards significance was also observed between
38 intercaruncular *SERPINA14* expression and caruncular *IFN* γ expression (sr: -0.771; P=
39 0.072). Luteal tissue *IL10* expression was positively correlated with *IFN* γ and *IL4*
40 expression both in luteal tissue (sr: 0.82; P=0.023 and sr: 0.714; P=0.071, respectively)
41 and in caruncle samples (sr: 0.83; P=0.042 and sr: 0.88; P=0.019, respectively). No
42 relationships were detected between cytokine gene expression at the foetal-maternal
43 interface and *SERPINA14* expression in the luteal samples. Our findings indicate that, in
44 these experimentally infected dams, gene expression of the uterine serpin *SERPINA14*
45 correlates negatively with the expression of Th1 and Th2 cytokines at the foetal-
46 maternal interface but not in the corpus luteum.

47

Keywords: Bovine neosporosis, pregnancy, maternal immune response, placenta, mummified foetus

Introduction

The mammalian foetus is an antigenically foreign body whose survival is the result of suppressed immunological interactions between mother and foetus (Druckmann 2001; Bauersachs and Wolf 2013). The key pregnancy hormone, progesterone, promotes mechanisms that induce this immunological tolerance of the foetus (Hansen et al. 1986). The immune response during pregnancy is mostly regulated through cytokines which are produced by T helper (Th) and T regulatory (Treg) cells. Cytokines are generally classified as “inflammatory cytokines” derived from Th1 cells and “pro-gestation cytokines” derived from Th2 cells. These two cytokine types stimulate cell-mediated immunity and promote the humoral response respectively (Mellor and Munn 2000). During the pregnancy period, progesterone seems to induce some Th2 bias, whereas an excessive Th1 response can induce pregnancy loss (Ragupathy 1997; Roberston 2000). Currently, it is not fully understood how the conceptus is able to avoid maternal immune attack. In addition, we would expect different conflict between reproductive and immune systems in healthy cows and those with a chronic infection.

Neospora caninum is an obligate, intracellular, protozoan parasite considered a major cause of abortion in cattle worldwide (Dubey and Schares 2011; Almería and López-Gatius 2015). On dairy farms, the major route of *N. caninum* infection is transplacental transmission from naturally infected dams to their foetuses during pregnancy (Goodswen et al. 2013; Almería and López-Gatius 2013). Most calves born to infected dams are clinically normal yet up to 95% of them remain infected for life (Dubey et al.

2007; Reichel et al. 2014). Abortion or congenital infection occurs when parasites cross the placenta and infect the foetus (Dubey and Lindsay 1996), and most abortions occur at 5-7 months of gestation (Almería and López-Gatius 2013). In our geographical area of study, northeastern Spain, *Neospora*-seropositive cows show a 12–19 times greater risk of abortion than seronegative cows, and the abortion rate ranges from 30 to 44% of seropositive animals (López-Gatius et al. 2004a,b). Foetuses may also die in utero and become mummified (Dubey and Lindsay 1996). The reasons why some animals abort and others do not remain unknown.

Th1 cytokines such as gamma interferon (IFN- γ) play an essential role in providing protective immunity against *N. caninum*. IFN- γ is a pro-inflammatory cytokine that inhibits the intracellular multiplication of *N. caninum* tachyzoites. However, an intense pro-inflammatory response, effective against *N. caninum* in non-pregnant cows, will be likely followed by foetal or placental damage and abortion in pregnant cows (Innes et al. 2005; Almeria and López-Gatius 2015). In cattle, IFN- γ has been linked to protection against *N. caninum*-associated abortion in several studies (Lopez-Gatius et al. 2007b; Williams et al. 2007; Almeria et al. 2012). Probably, the Th2 cytokine bias promotes maintenance of pregnancy by reducing local inflammatory responses (Wegmann et al. 1993; Chaouat et al. 2002). Our observation of the upregulated cytokine gene expression of both IFN γ and *IL4* (Th2) in infected dams reinforces this idea (Almería et al. 2016b).

It is known that *Neospora caninum* infection modifies endocrine patterns throughout gestation in dairy cattle. For example, *Neospora*-seropositivity has been associated with increased plasma prolactin and progesterone concentrations (Garcia-Ispuerto et al. 2009,

2010) and reduced plasma concentrations of pregnancy-associated glycoproteins (PAGs) (Garcia-Ispuerto et al. 2015). Pregnancy-associated glycoproteins I (PAG-I) and II (PAG-II) coexist in the ruminant trophoctoderm (Zoli et al. 1991; Garbayo et al. 2008). Although the functions of PAGs are not yet fully understood, PAG-I and PAG-II concentrations in aborting animals are useful indicators of foetal-placental impairment (López-Gatius et al. 2007a; Garcia-Ispuerto et al. 2013, 2105). In a recent study, expression patterns of the genes *SERPINA14*, *PAG1*, and *PAG2* at the foetal-maternal interface were investigated in dairy heifers experimentally infected with *N. caninum* during the second trimester of pregnancy (Serrano-Pérez et al. 2016). In infected dams with aborted fetuses, *SERPINA14* expression was significantly reduced and a negative relationship was observed between *N. caninum* antibody titres and *SERPINA14* or *PAG* expression in all infected animals (Serrano-Pérez et al. 2016).

The immunosuppressive actions of progesterone on the uterus during gestation have been attributed in part to uterine serpins (Hansen et al. 1987; Leslie and Hansen 1991). Serpins are glycoproteins and members of the serpin superfamily of serine peptidase inhibitors. One such serpin, SERPINA 14 (serpin peptidase inhibitor, clade A member 14), is expressed in response to progesterone in the endometrium (Ing and Roberts 1989) and has been linked to maternal immunosuppression during pregnancy (Padua and Hansen 2010). In pregnant ruminants, besides its presence in the endometrium, SERPINA14 also occurs in ovarian luteal and follicular structures (Ulbrich et al. 2009). Since *Neospora*-infection seems to reduce *SERPINA14* gene expression (Serrano-Pérez et al. 2016) and increase *IFN γ* and *IL4* gene expression (Almería et al. 2016b), we hypothesized that *SERPINA14* gene expression would be negatively correlated with the expression of *IFN γ* and *IL4* genes in pregnant dams suffering *Neospora*-infection.

123

124 The present study is one of a series of investigations performed during the second

125 trimester of gestation in pregnant dairy heifers experimentally infected with *N. caninum*

126 on day 110 of gestation. This stage of pregnancy was selected as the time when most

127 abortions occur in field conditions. The objective of this study was to detect possible

128 correlation between expression of the genes *IFN γ* , *IL4*, interleukin 10 (Treg) (*IL10*) and

129 *SERPINA14* at the foetal-maternal interface on day 152 of gestation in *Neospora*-

130 infected non-aborting animals. Since tropism of *N. caninum* for the ovarian follicle has

131 been suggested (Silva et al. 2012), a second objective was examine the same gene

132 expression patterns in the corpus luteum and their possible relationships with *Neospora*-

133 infection and with cytokine gene expression at the foetal-maternal interface.

134

135 Material and methods

136

137 Animals and infection

138

139 The animals used and the infection protocol have been described elsewhere (Almeria et

140 al. 2016a). Briefly, ten 14-16 month-old Holstein-Friesian heifers free of abortifacient

141 agents and seronegative for *N. caninum* (CIVTEST® anti-*Neospora*; Hipra, Girona,

142 Spain) were synchronized for oestrus and artificially inseminated. Seronegativity was

143 confirmed before insemination and on Days 60, 90 and 110 of gestation. Pregnancy was

144 assessed by ultrasonography 30, 45, 90 and 110 days after insemination. On Day 110 of

145 gestation, six of the heifers were intravenously (i.v.) inoculated with 10⁷ culture-derived

146 tachyzoites of the *N. caninum* isolate Nc-Spain7, kindly donated by Dr. L. M. Ortega-

147 Mora (SALUVET, Universidad Complutense, Madrid, Spain). Two heifers that aborted

1
2
3 148 at 14 and 21 days post-infection were excluded from the study. Another heifer had a
4
5 149 mummified foetus upon euthanasia. The four non-aborting animals were euthanized on
6
7 150 Day 152 of gestation. The four remaining heifers were kept as un-inoculated controls
8
9 151 and were euthanized at the same time as the inoculated dams.
10
11
12 152

13 153 Sample collection

14
15
16 154

17
18 155 On Day 152 of gestation (Day 42 post-infection), blood samples were collected by tail
19
20 156 vein puncture into heparinized vacuum tubes (BD Vacutainer, Becton-Dickinson and
21
22 157 Company, Plymouth, UK) to determine maternal antibodies and progesterone
23
24 158 concentrations. Plasma obtained by centrifugation within 30 min of sampling was stored
25
26 159 at -20°C until analysis. After blood collection, all animals were sedated with xylazine
27
28 160 hydrochloride (Rompun; Bayer, Sant Joan Despí, Barcelona, Spain) and immediately
29
30 161 euthanized by an intravenous (i.v.) overdose of embutramide and mebezonio iodide
31
32 162 (T61; Intervet, Salamanca, Spain). Immediately after sacrifice, heifers were necropsied
33
34 163 and tissues were removed aseptically according to Almería et al. (2016a). The uterus
35
36 164 was removed and foetal amniotic and allantoic fluids collected by puncture (Mur-
37
38 165 Novales et al. 2016). Next, the uterus was opened and foetal tissue and blood samples
39
40 166 were collected. Portions of foetal tissues were aseptically obtained and stored in liquid
41
42 167 nitrogen at -196°C until DNA extraction. Tissues collected from foetuses were: CNS
43
44 168 (brain and spinal cord), heart, lung, liver, skeletal muscle, spleen, and thymus. The
45
46 169 lymphatic vessels draining the uterus, or internal iliac lymph nodes referred to here as
47
48 170 uterine lymph nodes (UTLN), were collected from the heifers for the isolation of
49
50 171 mononuclear cells as described by Almería et al. (2014). In addition, samples of three
51
52 172 selected placentomes (cranial, medial, and caudal placenta) were removed from each
53
54
55
56
57
58
59
60

173 dam. Both the maternal side (caruncle) and foetal side of the placenta (cotyledon) were
174 careful separated manually from each placentome. Intercaruncular tissue was also
175 collected. The corpora lutea were aseptically dissected and divided into two sections:
176 one was stored in liquid nitrogen and the other section was used for histopathology.

178 Ethics

180 All procedures were approved by the Ethics Committees on Animal Experimentation of
181 the Autonomous University of Barcelona (license number CEEAH.1426-08/02/2012)
182 and University of Lleida (license number CEEA.06-01/12). Animals were handled in
183 strict accordance with good animal practices and the conditions defined by the Animal
184 Ethics Committee of the Autonomous University of Barcelona and CReSA, Spain.
185 Every effort was made to minimize suffering.

187 Sample analysis

189 Progesterone assay and corpus luteum histopathology

191 Progesterone concentrations were determined in plasma samples using an ELISA kit
192 designed for bovine plasma (Ridgeway Science, St. Briavels, Gloucestershire, UK),
193 according to the manufacturer's instructions. Assay sensitivity was 0.33 ng/mL. All
194 samples were analysed in duplicate in the same assay. Intra-assay coefficients of
195 variation for sample pools of 1, and 2 ng/mL were 6.4 and 4.8%, respectively.

197 Paraffin-embedded 5- μ m sections of corpora lutea were prepared and stained with
198 haematoxylin-eosin for histopathological examinations.

199

200 Total RNA extraction and cDNA synthesis

201

202 Intercaruncular, UTLN, placental and luteal tissue samples were kept frozen in liquid
203 nitrogen, homogenized in a mortar in the presence of additional liquid nitrogen and
204 maintained in trizol (Invitrogen Corp., Carlsbad, CA, USA) at -80°C. For caruncle or
205 cotyledon tissue gene expression analysis, a mixed sample of RNA from the three
206 different sections (cranial, medial and caudal) of each tissue was used as template.

207

208 Total RNA was extracted according to the method of Chomczynski and Sacchi (1987).

209 Samples were treated with DNase in the presence of RNase inhibitors to eliminate

210 contaminating genomic DNA. Concentrations of RNA were determined

211 spectrophotometrically. RNA integrity was checked by denaturing agarose gel

212 electrophoresis. Complementary DNA was synthesized from 2 μ g of total RNA in the

213 presence of random primers using the High Capacity cDNA Reverse Transcription kit

214 (Life Technologies, Carlsbad, CA, USA) according to the manufacturer's

215 recommendations.

216

217 Real time RT-PCR

218

219 Messenger RNA expression was determined by real time RT-PCR on four target genes:

220 *IFN γ* , *IL4*, *IL10* and *SERPINA14*.

221

222 Cytokine gene expression

223

224 Cytokine gene expression (*IFN γ* , *IL4*, *IL10*) was determined in UTLN, placental and

225 luteal samples. Messenger RNA expression was determined by real time RT-PCR

226 following the Taqman approach in an ABI PRISMTM 7700 sequence detector (PE

227 Applied Biosystem, Foster city, CA, USA). Probes and primers for bovine *IL4* and *IFN γ*

228 were those reported by Waldvogel et al. (2000). The bovine *IL10* sequence probes and

229 primers used have been described in Almeria et al. (2003). Probes and primer pairs were

230 used to quantify *GAPDH* RNA as the endogenous housekeeping control gene described

231 by Leutenegger et al. (2000). Primer and probe concentrations for each cytokine were

232 determined as previously reported (Almeria et al. 2003, 2011). PCR amplifications were

233 conducted as in Almeria et al. (2014). Endogenous *GAPDH* housekeeping expression

234 was used to normalize levels of cytokine gene expression.

235

236 *SERPINA14* gene expression

237

238 *SERPINA14* gene expression was determined by real-time PCR in intercaruncular,

239 placental and luteal samples. *SERPINA14* mRNA expression was determined using the

240 SYBRgreen method and ABI PRISMTM 7500 sequence detector (Applied Biosystem,

241 Foster City, CA, USA). The gene β -actin (*ACTB*) was used as housekeeping gene to

242 normalize levels of *SERPINA14* gene expression. The primers used for *ACTB* and

243 *SERPINA14* have been described elsewhere (Ribeiro et al. 2014; Serrano-Pérez et al.

244 2016). Amplifications were performed as described in Serrano-Pérez et al. (2016).

245

For relative quantification of gene expression, the comparative threshold cycle (CT) method (ABI PRISM7700 sequence detection system, user bulletin #2) was used as described in Almeria et al. (2003). Briefly, the CT value for the housekeeping gene was subtracted from the CT value for each target expression gene to normalize RNA content and provide a relative expression value for each target gene. This value was defined as ΔCT . To assess the effects of infection, the mean ΔCT for the control dams was subtracted from the mean ΔCT determined for the *Neospora*-infected dams. This value was defined as $\Delta\Delta CT$. Relative fold increases or decreases were then calculated as $2^{-\Delta\Delta CT}$.

Statistical analysis

Spearman's rho (sr) test was used to identify possible relationships between gene expression levels of each cytokine and *SERPINA14* in uninfected controls and infected animals. The Student's t-test was used to compare relative *SERPINA14* and cytokine mRNA expression in luteal samples and progesterone concentrations among uninfected controls and infected dams. Only dams with viable foetus upon euthanasia were included in the latter statistical analysis. All tests were performed using the computer package SPSS version 17.0 (SPSS Inc., Chicago, IL). Significance was set at $P \leq 0.05$.

Results

The present study was designed on the basis of the results described in Almería et al. (2016). In this prior work, all experimentally infected heifers were seropositive for *N. caninum* upon euthanasia, and transplacental infection had already taken place in their

foetuses. Control uninfected foetuses showed no antibodies and *N. caninum* DNA was not detected in any of their tissues. The mummified foetus was estimated to have died approximately 28 days after infection (Almeria et al. 2016a). Due to the lack or poor condition of some samples, a variable number of samples for cytokine and *SERPINA14* gene expression were available: samples from seven animals for UTLN and caruncle tissues (controls, n=3; infected dams, n=4), samples from eight animals for intercaruncular and luteal tissues (controls, n=4; infected dams=4), and samples from six animals for cotyledon tissues (controls, n=3; infected dams=3).

Gene expression levels in the dam with the mummified foetus were similar to those in the remaining three infected dams in the intercaruncular (*SERPINA14*), UTLN (*IFN γ* , *IL4*, *IL10*) and luteal tissues (*IFN γ* , *IL4*, *IL10*, *SERPINA14*).

Plasma progesterone determinations and luteal histopathology

Neospora-infection did not affect plasma progesterone concentrations (mean \pm S.D. values: 12.53 ± 1.5 ng/ml in controls vs. 12.85 ± 3.7 ng/ml in infected dams) and microscopic lesions were not observed in the luteal samples of any of the 4 infected dams.

Correlations between *SERPINA14* and cytokine gene expression in UTLN, intercaruncular and placental tissues

SERPINA14 expression in intercaruncular samples was negatively correlated with *IFN γ* , expression in cotyledon samples (sr: -0.943; P= 0.005) and with *IL4* expression in

UTLN samples (sr: -0.886; P= 0.019). Trends towards significance were also observed between *SERPINA14* expression in intercaruncular samples and *IFN γ* in caruncle samples (sr: -0.771; P= 0.072). No other significant correlations were found.

Gene expression of *SERPINA14* and cytokines in luteal samples

Luteal samples from uninfected heifers did not reach detection threshold levels of expression for *IL4*. When normalized levels were compared, no relationships were detected between any cytokine and *SERPINA14* gene expression. In addition, no significant differences were detected between the gene expression of cytokines and *SERPINA14* in luteal samples from infected dams and control dams (Figure 1). Cytokine and *SERPINA 14* gene expression levels detected in the infected dam with a mummified foetus upon euthanasia, not included in our statistical analysis, are also shown in Figure 1.

Correlating *SERPINA14* and cytokine gene expression between luteal and foetal-maternal interface samples

The expression of *IL10* in luteal samples was positively correlated with *IFN γ* and *IL4* expression in both luteal samples (sr: 0.82; P=0.023 and sr: 0.714; P=0.071, respectively) and caruncle samples (sr: 0.83; P=0.042 and sr: 0.88; P=0.019, respectively). No relationships were detected between any cytokine expressed at the foetal-maternal interface and *SERPINA14* gene expression in the luteal samples.

Discussion

321

322 This study unveils several aspects of immune-reproductive modulation at the level of

323 the foetal-maternal interface in response to *N. caninum* infection during the second

324 trimester of gestation in dairy heifers. Relationships between patterns of gene

325 expression shown by a uterine serpin (*SERPINA14*) and by the cytokines Th1 (*IFN γ*),

326 Th2 (*IL4*) and Treg (*IL10*) in UTLN, intercaruncular, placental and luteal tissues

327 provided useful insight into this modulation. The most noteworthy findings of the

328 present study were that *SERPINA14* gene expression in intercaruncular tissues

329 correlated negatively with *IFN γ* gene expression in the cotyledons and with *IL4* in the

330 UTLN tissue samples, whilst *IL10* expression in corpus luteum correlated positively

331 with *IFN γ* and *IL4* in both luteal and placental tissues.

332

333 Immunosuppressive function downregulation by SERPINA 14 in response to *N.*

334 *caninum* infection was suggested in a prior study (Serrano-Pérez et al. 2016). In effect,

335 the negative correlation observed here between *SERPINA14* expression and Th1 and

336 Th2 cytokines likely indicates that reduced local immunosuppression by the uterine

337 serpin is needed to improve maternal immune responses against the parasite to maintain

338 gestation. These data support recent findings involving the production *in vitro* of the

339 cytokines IFN- γ (Th1) and IL-4 (Th2) whereby a protective immune response against

340 abortion could not be associated with IFN- γ levels alone, but was significantly linked to

341 lower IFN- γ /IL-4 ratios (Darwich et al. 2016). Thus, the Th1/Th2 balance seems to play

342 a key role in maintaining pregnancy during the course of *N. caninum* infection.

343

344 Our observation of significant positive correlation between the luteal expression of Treg

345 (*IL10*) and the expression of Th1 (*IFN γ*) and Th2 (*IL4*) in both luteal and placental

1
2
3 346 tissues is consistent with several reports of simultaneously up-regulated expression of
4
5 347 the different cytokine subsets in pregnant cattle at the placental level during *N. caninum*
6
7 348 infection (Rosbottom et al., 2008; Almeria et al. 2011, 2014, 2016b; Cantón et al. 2014;
8
9 349 Regidor-Cerrillo et al., 2014; Hecker et al. 2015). Thus, it seems the placenta can act as
10
11 350 an immune organ that can sense changes in the environment and then signal to other
12
13 351 cells to elicit a response (Schminkey and Groer 2014; PrabhuDas et al. 2015). The
14
15 352 currently held type 1/type 2 paradigm of pregnancy indicates that to maintain uterine
16
17 353 quiescence, Th2 cytokines predominate throughout mid-pregnancy, while the beginning
18
19 354 and end of pregnancy are times of type 1 predominance allowing for implantation and
20
21 355 parturition. This mechanism could be controlled by the innate immune system, which
22
23 356 responds to the pregnancy in much the same way as it responds to other acute or
24
25 357 naturalistic stressors (Schminkey and Groer, 2014).
26
27
28
29
30

31
32 359 While *SERPINA14* was observed to correlate negatively with cytokine production at the
33
34 360 foetal-maternal interface, no relationships were detected between any cytokine and
35
36 361 *SERPINA14* gene expression in luteal samples. In addition, despite suggestions of
37
38 362 possible tropism of *N. caninum* for ovarian follicles (Silva et al. 2012), in our
39
40 363 experimental conditions, *N. caninum* infection had no clear effects on the immune
41
42 364 response taking place in the corpus luteum. Therefore, it seems that the infection had no
43
44 365 impacts on corpus luteal function, at least in terms of repercussions on plasma
45
46 366 progesterone concentrations and expression of the genes coding for *SERPINA14* and the
47
48 367 cytokines *IFN γ* , *IL4* and *IL10*.
49
50
51

52 368

53
54 369 In conclusion, our findings indicate that the gene expression of the uterine serpin
55
56 370 *SERPINA14* correlates negatively with the expression of Th1 and Th2 cytokines at the
57
58
59
60

371 foetal-maternal interface but not the corpus luteum in dams experimentally infected
372 with *N. caninum* during the second trimester of gestation. These data are consistent with
373 our starting hypothesis that expression of the *SERPINA14* gene would correlate
374 negatively with the expression of the *IFN γ* and *IL4* genes.

376 Acknowledgements

378 This study was supported by a grant from the Spanish MINECO (AGL2012-39830-
379 C02-01/02) and FEDER. The authors thank Ana Burton for editorial assistance.

381 References

- 383 Almería S, López-Gatius F, 2013: Bovine neosporosis: clinical and practical aspects.
384 Res Vet Sci **95**, 303-309.
- 386 Almería S, López-Gatius F, 2015: Markers related to the diagnosis and to the risk of
387 abortion in bovine neosporosis. Res Vet Sci **100**, 169-175.
- 389 Almería S, Ferrer D, Pabón M, Castellà J, Manas S, 2002: Red foxes (*Vulpes vulpes*)
390 are natural intermediate host of *Neospora caninum*. Vet Parasitol **107**, 287–294.
- 392 Almería S, De Marez T, Dawson H, Araujo R, Dubey JP, Gasbarre LC, 2003: Cytokine
393 gene expression in dams and foetuses after experimental *Neospora caninum* infection of
394 heifers at 110 days of gestation. Parasite Immunol **25**, 383-392.

- 395
- 396 Almería S, Araujo RN, Darwich L, Dubey JP, Gasbarre LC, 2011: Cytokine gene
397 expression at the materno-foetal interface after experimental *Neospora caninum*
398 infection of heifers at 110 days of gestation. *Parasite Immunol* **33**, 517-523.
399
- 400 Almería S, Serrano B, Yàñez JL, Darwich L, López-Gatius F, 2012: Cytokine gene
401 expression profiles in peripheral blood mononuclear cells from *Neospora caninum*
402 naturally infected dams throughout gestation. *Vet Parasitol* **183**, 237-243.
403
- 404 Almería S, Serrano-Pérez B, Darwich L, Araujo RN, Lopez-Gatius F, Dubey JP,
405 Gasbarre LC, 2014: Maternal and foetal immune response patterns in heifers
406 experimentally infected with *Neospora caninum* in the second trimester of pregnancy-a
407 descriptive study. *Vet Parasitol* **204**, 146-152.
408
- 409 Almería S, Serrano-Pérez B, Darwich L, Domingo M, Mur-Novales R, Regidor-Cerrillo
410 J, Cabezón O, Pérez-Maillo M, López-Helguera I, Fernández-Aguilar X, Puig-Ribas M,
411 Ortega-Mora LM, García-Ispuerto I, Dubey JP, López-Gatius F, 2016a: Foetal death in
412 naive heifers inoculated with *Neospora caninum* isolate Nc-Spain7 at 110 days of
413 pregnancy. *Exp Parasitol* **168**, 62-69.
414
- 415 Almería S, Serrano-Pérez B, Darwich L, Mur-Novales R, García-Ispuerto I, Cabezón O,
416 López-Gatius F, 2016b: Cytokine gene expression in aborting and non-aborting dams
417 and in their foetuses after experimental infection with *Neospora caninum* at 110 days of
418 gestation. *Vet Parasitol* **227**, 138-142.
419

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

420 Bauersachs S, Wolf E, 2013: Immune aspects of embryo-maternal cross-talk in the
421 bovine uterus. J Reprod Immunol **97**, 20-26.
422
423 Cantón GJ, Katzer F, Maley SW, Bartley PM, Benavides-Silván J, Palarea-Albaladejo J,
424 Pang Y, Smith SH, Rocchi M, Buxton D, Innes EA, Chianini F, 2014: Cytokine
425 expression in the placenta of pregnant cattle after inoculation with *Neospora caninum*.
426 Vet Immunol Immunopathol **161**, 77-89.
427
428 Chaouat G, Zourbas S, Ostojic S, Lappree-Delage G, Dubanchet S, Ledee N, Martal J,
429 2002: A brief review of recent data on some cytokine expressions at the materno-foetal
430 interface which might challenge the classical Th1/Th2 dichotomy. J Reprod Immunol.
431 53, 241–256.
432
433 Chomczynski P, Sacchi N, 1987: Single-step method of RNA isolation by acid
434 guanidinium thiocyanate phenol chloroform extraction. Anal Biochem 162, 156-159.
435
436 Darwich L, Li Y, Serrano-Pérez B, Mur-Novales R, Garcia-Ispuerto I, Cabezón O,
437 López-Gatius F, Almería S, 2016: Maternal and foetal cytokine production in dams
438 naturally and experimentally infected with *Neospora caninum* on gestation Day 110.
439 Res Vet Sci **107**, 55-61.
440
441 Druckmann R, 2001: Review: female sex hormones, autoimmune diseases and immune
442 response. Gynecol Endocrinol **5(Suppl 6)**, 69-76.
443
444 Dubey JP, Schares G, 2006: Diagnosis of bovine neosporosis. Parasitol **140**, 1-34.

- 445
- 446 Dubey JP, Lindsay DS, 1996: A review of *Neospora caninum* and neosporosis. Vet
- 447 Parasitol **67**, 1-59.
- 448
- 449 Dubey JP, Schares G, Ortega-Mora LM, 2007: Epidemiology and control of
- 450 neosporosis and *Neospora caninum*. Clin Microbiol Rev **20**, 323-367.
- 451
- 452 Garbayo JM, Serrano B, Lopez-Gatius F, 2008: Identification of novel pregnancy-
- 453 associated glycoproteins (PAG) expressed by the peri-implantation conceptus of
- 454 domestic ruminants. Anim Reprod Sci **103**, 120–134.
- 455
- 456 García-Ispuerto I, Nogareda C, Yániz JL, Almería S, Martínez-Bello D, de Sousa NM,
- 457 Beckers JF, López-Gatius F, 2010: *Neospora caninum* and *Coxiella burnetii*
- 458 seropositivity are related to endocrine pattern changes during gestation in lactating dairy
- 459 cows. Theriogenology **74**, 212–220.
- 460
- 461 García-Ispuerto I, Almería S, Serrano B, de Sousa NM, Beckers JF, López-Gatius F,
- 462 2013: Plasma concentrations of pregnancy-associated glycoproteins measured using
- 463 anti-bovine PAG-2 antibodies on day 120 of gestation predict abortion in dairy cows
- 464 naturally infected with *Neospora caninum*. Reprod Domest Anim **48**, 613-618.
- 465
- 466 García-Ispuerto I, Serrano-Pérez B, Almería S, Martínez-Bello D, Tchimbou AF, de
- 467 Sousa NM, Beckers JF, López-Gatius F, 2015: Effects of crossbreeding on endocrine
- 468 patterns determined in pregnant beef/dairy cows naturally infected with *Neospora*
- 469 *caninum*. Theriogenology **83**, 491–496.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

470

471 Goodswen SJ, Kennedy PJ, Ellis JT, 2013: A review of the infection, genetics, and

472 evolution of *Neospora caninum*: from the past to the present. Infect Genet Evol **13**, 133-

473 150.

474

475 Hansen PJ, Bazer FW, Segerson EC Jr, 1986: Skin graft survival in the uterine lumen of

476 ewes treated with progesterone. Am J Reprod Immunol Microbiol 1986 **12**, 48-54.

477

478 Hansen PJ, Ing NH, Moffatt RJ, Baumbach GA, Saunders PT, Bazer FW, Roberts RM,

479 1987: Biochemical characterization and biosynthesis of the uterine milk proteins of the

480 pregnant sheep uterus. Biol Reprod **36**, 405-418.

481

482 Hecker YP, Cantón G, Regidor-Cerrillo J, Chianini F, Morrell E, Lischinsky L, Ortega-

483 Mora LM, Innes EA, Odeón A, Campero CM, Moore DP, 2015: Cell mediated immune

484 responses in the placenta following challenge of vaccinated pregnant heifers with

485 *Neospora caninum*. Vet Parasitol **214**, 247-254.

486

487 Ing NH, Roberts RM, 1989: The major progesterone-modulated proteins secreted into

488 the sheep uterus are members of the serpin superfamily of serine protease inhibitors. J

489 Biol Chem 264, 3372-3379.

490

491 Innes EA, Wright S, Bartley P, Maley S, Macaldowie C, Esteban-Redondo I, Buxton D,

492 2005: The host-parasite relationship in bovine neosporosis. Vet Immunol Immunopathol

493 **108**, 29-36.

494

- 495 Leslie MV, Hansen PJ, 1991: Progesterone-regulated secretion of the serpin-like
496 proteins of the ovine and bovine uterus. *Steroids* 56, 589-597.
497
- 498 Leutenegger CM, Alluwaimi AM, Smith WL, Perani L, Cullor JS, 2000: Quantitation of
499 bovine cytokine mRNA in milk cells of healthy cattle by real-time TaqMan polymerase
500 chain reaction. *Vet Immunol Immunopathol.* 77, 275-287.
501
- 502 López-Gatius F, López-Béjar M, Murugavel K, Pabón M, Ferrer D, Almería S, 2004a:
503 *Neospora*-associated abortion episode over a 1-year period in a dairy herd in north-east
504 Spain. *J Vet Med B* 51, 348-352.
505
- 506 López-Gatius F, Pabon M, Almería S, 2004b: *Neospora caninum* infection does not
507 affect early pregnancy in dairy cattle. *Theriogenology* 62, 606-613.
508
- 509 López-Gatius F, Garbayo JM, Santolaria P, Yániz JL, Almería S, Ayad A, de Sousa
510 NM, Beckers JF, 2007a: Plasma pregnancy- associated glycoprotein-1 (PAG-1)
511 concentrations during gestation in *Neospora*- infected dairy cows. *Theriogenology* 67,
512 502-508.
513
- 514 López-Gatius F, Almería S, Donofrio G, Nogareda C, García-Ispuerto I, Bech-Sabat G,
515 Santolaria P, Yaniz JL, Pabón M, de Sousa NM, Beckers JF, 2007b: Protection against
516 abortion linked to gamma interferon production in pregnant dairy cows naturally
517 infected with *Neospora caninum*. *Theriogenology* 68, 1067-1073.
518

1
2
3 519 Mellor AL, Munn DH, 2000: Immunology at the maternal-fetal interface: lessons for T
4
5 520 cell tolerance and suppression. *Annu Rev Immunol* **18**, 367-391.
6
7 521
8
9 522 Mur-Novales R, López-Gatius F, Serrano-Pérez B, García-Ispuerto I, Darwich L,
10
11 523 Cabezón O, de Sousa NM, Beckers JF, Almería S, 2016: Experimental *Neospora*
12
13 524 *caninum* infection in pregnant dairy heifers raises concentrations of pregnancy-
14
15 525 associated glycoproteins 1 and 2 in foetal fluids. *Reprod Domest Anim* **51**, 282-286.
16
17 526
18
19
20 527 PrabhuDas M, Bonney E, Caron K, Dey S, Erlebacher A, Fazleabas A, Fisher S, Golos
21
22 528 T, Matzuk M, McCune JM, Mor G, Schulz L, Soares M, Spencer T, Strominger J, Way
23
24 529 SS, Yoshinaga K, 2015: Immune mechanisms at the maternal-fetal interface:
25
26 530 perspectives and challenges. *Nature Immunol* **16**, 328-334.
27
28 531
29
30 532 Raghupathy R, 1997: Th-1 immunity is incompatible with successful pregnancy.
31
32 533 *Immunol Today* **18**, 478-482.
33
34 534
35
36 535 Regidor-Cerrillo J, Arranz-Solís D, Benavides J, Gómez-Bautista M, Castro-Hermida
37
38 536 JA, Mezo M, Pérez V, Ortega-Mora LM, González-Warleta M, 2014: *Neospora*
39
40 537 *caninum* infection during early pregnancy in cattle: how the isolate influences infection
41
42 538 dynamics, clinical outcome and peripheral and local immune responses. *Vet Res* **45**, 10.
43
44 539
45
46 540 Reichel MP, McAllister MM, Pomroy WE, Campero C, Ortega-Mora LM, Ellis JT,
47
48 541 2014: Control options for *Neospora caninum*-is there anything new or are we going
49
50 542 backwards? *Parasitol* **141**, 1455-1470.
51
52 543
53
54
55
56
57
58
59
60

- 1
2
3 544 Ribeiro ES, Bruno RG, Farias AM, Hernández-Rivera JA, Gomes GC, Surjus R, Becker
4
5 545 LF, Birt A, Ott TL, Branen JR, Sasser RG, Keisler DH, Thatcher WW, Bilby TR,
6
7 546 Santos JEP, 2014: Low doses of bovine somatotropin enhance conceptus development
8
9 547 and fertility in lactating dairy cows. Biol Reprod **90(1):10**, 1-12.
10
11 548
12
13 549 Roberston SA, 2000: Control of the immunological environment of the uterus. Rev
14
15 550 Reprod **5**, 164-164.
16
17 551
18
19 552 Rosbottom A, Gibney EH, Guy CS, Kipar A, Smith RF, Kaiser P, Trees AJ, Williams
20
21 553 DJL, 2008: Upregulation of cytokines is detected in the placentas of cattle infected with
22
23 554 *Neospora caninum* and is more marked early in gestation when foetal death is observed.
24
25 555 Infect Immun **76**, 2352-2361.
26
27 556
28
29 557 Schminkey DL, Groer M, 2014: Imitating a stress response: A new hypothesis about
30
31 558 the innate immune system's role in pregnancy. Med Hypotheses **82**, 721–729.
32
33 559
34
35 560 Serrano-Pérez B, Hansen PJ, Mur-Novales R, Garcia-Ispuerto I, de Sousa NM, Beckers
36
37 561 JF, Almería S, López-Gatius F, 2016: Crosstalk between uterine serpin (SERPINA 14)
38
39 562 and pregnancy-associated glycoproteins at the fetal-maternal interface in pregnant dairy
40
41 563 heifers experimentally infected with *Neospora caninum*. Theriogenology **86**, 824-830.
42
43 564
44
45 565 Silva AF, Rangel L, Ortiz CG, Morales E, Zanella EL, Castillo-Velázquez U, Gutiérrez
46
47 566 CG, 2012: Increased incidence of DNA amplification in follicular than in uterine and
48
49 567 blood samples indicates possible tropism of *Neospora caninum* to ovarian follicle. Vet
50
51 568 Parasitol **188**, 175-178.
52
53
54
55
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

569

570 Ulbrich SE, Frohlich T, Schulke K, Englberger E, Waldschmitt N, Arnold GJ,
571 Reichenbach HD, Reichenbach M, Wolf E, Meyer HH, Bauersachs S, 2009: Evidence
572 for estrogen-dependent uterine serpin (SERPINA14) expression during estrus in the
573 bovine endometrial glandular epithelium and lumen. Biol Reprod **81**, 795-805.

574

575 Waldvogel AS, Hediger-Weithaler BM, Eicher R, Zakher A, Zarlenga DS, Gasbarre
576 LC, Heussler VT, 2000: Interferon- γ and interleukin-4 mRNA expression by peripheral
577 blood mononuclear cells from pregnant and non-pregnant cattle seropositive for bovine
578 viral diarrhea virus. Vet Immunol Immunopathol **77**, 201–212.

579

580 Wegmann, TG, Lin H, Guilbert L, Mosmann TR, 1993: Bidirectional cytokine
581 interactions in the maternal-foetal relationship: is successful pregnancy a TH2
582 phenomenon? Immunol Today **14**, 353–356.

583

584 Williams DJ, Guy CS, Smith RF, Ellis J, Björkman C, Reichel MP, Trees AJ, 2007:
585 Immunization of cattle with live tachyzoites of *Neospora caninum* confers protection
586 against foetal death. Infect Immun **75**, 1343-1348.

Figure 1. Relative expression levels of the genes *SERPINA14*, *IFN γ* , *IL4* and *IL10* in luteal samples from control dams (n=4), *Neospora*-infected dams with viable foetuses (n=3), and dams with a mummified foetus (n=1) on Day 152 of gestation upon euthanasia. The higher the normalized CT value the lower the expression level. Bars represent mean Ct values \pm standard error of the mean.

For Peer Review